

Exercise and immune system as modulators of intestinal microbiome: implications for the gut-muscle axis hypothesis

Andrea Ticinesi^{1,2,3}, Fulvio Lauretani^{2*}, Claudio Tana², Antonio Nouvenne^{2,3}, Erminia Ridolo^{1,2}, Tiziana Meschi^{1,2,3}

¹ Department of Medicine and Surgery, University of Parma, Italy

² Geriatric-Rehabilitation Department, Azienda Ospedaliero-Universitaria di Parma, Italy

³ Microbiome Research Hub, University of Parma, Italy

ABSTRACT

Exercise is a possible modulator of intestinal microbiome composition, since some investigations have shown that it is associated with increased biodiversity and representation of taxa with beneficial metabolic functions. Conversely, training to exhaustion can be associated with dysbiosis of the intestinal microbiome, promoting inflammation and negative metabolic consequences. Gut microbiota can, in turn, influence the pathophysiology of several distant organs, including the skeletal muscle. A gut-muscle axis may in fact regulate muscle protein deposition and muscle function. In older individuals, this axis may be involved in the pathogenesis of muscle wasting disorders through multiple mechanisms, involving transduction of pro-anabolic stimuli from dietary nutrients, modulation of inflammation and insulin sensitivity. The immune system plays a fundamental role in these processes, being influenced by microbiome composition and at the same time contributing to shape microbial communities. In this review, we summarize the most recent literature acquisitions in this field, disentangling the complex relationships between exercise, microbiome, immune system and skeletal muscle function and proposing an interpretative framework that will need verification in future studies.

Keywords: Gut microbiota; Sarcopenia; Inflammation; Exercise immunology; Sport

*Corresponding author:

Dr. Fulvio Lauretani, Department of Medicine and Surgery, University of Parma, Geriatric-Rehabilitation Department, Azienda Ospedaliero-Universitaria di Parma
Via Antonio Gramsci 14, 43126 Parma (Italy)
Tel: +39 3470912031, E-mail: flauratani@ao.pr.it

1. Introduction

1.1 The physiology of human intestinal microbiome

The human intestinal microbiome is composed of a complex ecosystem of more than 10^{14} bacteria, viruses, fungi, Protozoa and Archea that live symbiotically with the host in the gut lumen (45,61,82). Although there is increasing interest in the role of Protozoa, fungi (the “mycome”) and viruses (the “virome”), most of the existing research has been focused mainly on bacteria, thanks to the availability of high-throughput sequencing techniques of bacterial DNA (16S rRNA microbial profiling, shotgun metagenomics) and fecal metabolomics (52,77,93,94).

The concentration of bacteria generally increases from the small intestine to the colon, and different bacterial populations are harbored in different tracts of the gastrointestinal system (123). Although recent research has shown that the fecal microbiota composition does not completely overlap with the intestinal mucosa-associated microbiota (134), gut microbiota composition has been determined from fecal samples in most studies (45,61,82,123).

It is generally agreed that the intestinal microbiome composition is shaped during early childhood, influenced by genetic and environmental factors. These factors include geography, delivery mode, breastfeeding, weaning, and exposure to environmental bacteria (34,54,78). The maturation of the gut microbiota towards the adult-type is reached by the age of 3 years (128). Interestingly, this process seems to exert a great influence on the development of the immune system, promoting immunogenic tolerance towards symbionts and immune activation against pathogens (40,50,110). Conversely, alterations in this process may induce a shift towards unappropriated type 2 immune responses, favoring the pathogenesis of allergy or autoimmune diseases, according to some theories (71,74).

In adulthood, the human gut microbiota composition remains relatively stable over time, exhibiting resilience to disruptors, such as stress, acute diseases, or antibiotic administration (65). This means that, after a brief exposure to disruptors, the microbiome faces a substantial, but transient, perturbation, followed by partial or total recovery of previous taxonomic composition (96).

The healthy microbiome includes a limited number of highly represented taxa, such as *Bacteroides* and *Prevotella* spp., and a large number (up to 2000 identified to date) of minor players with low representation but high metabolic activity (42,65). In

healthy subjects, microbiota composition shows a certain degree of inter-individual differences, that can be again explained by genetic (13) and environmental factors (28), such as the place of living, cultural habits, diet, diseases, medications, and immune system function (99). Dietary factors, and particularly carbohydrate and protein intake, are generally considered as the main determinants of gut microbiota composition (5,76,132).

In the older age, species richness and diversity of microbiome decline, inter-individual variability increases, and resilience to perturbations is reduced (24,86). Thus, the microbiome composition shifts towards a higher representation of opportunistic pathogens, and is related to increasing prevalence of malnutrition, frailty, disability and multimorbidity (49,112,114).

1.2 The intestinal microbiome in human diseases

A large body of literature has demonstrated that the presence of acute and chronic diseases, not limited only to the gastrointestinal system, is associated with alterations of the gut microbiota composition (67,99). These alterations, globally referred to as “dysbiosis”, include reduced biodiversity, loss of commensals with possible beneficial metabolic activities and overgrowth of opportunistic pathogens (55,60,81). Dysbiosis implies a disruption of the mutual equilibrium between gut bacteria and host physiology (16). As a consequence, intestinal permeability increases, allowing bacteria or bacterial toxins and metabolites to enter into the host circulation and promote subclinical inflammation (64). Dysbiosis also reduces bioavailability of nutrients, affecting the microbial metabolism of several beneficial substances (73).

As a result, gut microbiota may exert a great influence on the functionality and pathophysiology of several organs anatomically distant from the gastrointestinal tract. For example, gut microbiota dysbiosis may be involved in the pathophysiology of dementia (17,115), Parkinson’s disease (108) (“gut-brain axis”), chronic kidney disease (95), nephrolithiasis (113) (“gut-kidney axis”), asthma (73) (“gut-lung axis”) and even osteoporosis (84) (“gut-bone axis”).

However, most of these associations linking dysbiosis with extra-intestinal diseases have been demonstrated in studies with a cross-sectional design. Longitudinal studies, demonstrating a causal relationship between microbiota composition and disease onset and course, are still lacking (99). Thus, the real clinical implications of gut microbiota dysbiosis and interventions targeted at modifying gut microbiota composition are still poorly understood.

2. Exercise and gut microbiota: the yin and yang

2.1 Beneficial effects of exercise on gut microbiota

Exercise is considered as one of the main environmental factors possibly influencing gut microbiota composition (99). The complex relationship between exercise and microbiome, and its possible implications for athletic performance have already been reviewed elsewhere (10,27,87). However, research in this field has made substantial improvements in the last few years, and some recent acquisitions deserve to be mentioned and discussed.

Exercise is generally considered a positive modulator of gut microbiota biodiversity. This concept has been supported by investigations performed in animals (8,18,57,75), and then confirmed in human studies.

In a case-control study, microbial diversity was much higher in a group of professional rugby players than in age-, sex- and body size-matched controls not performing sports (26). Recently, shotgun metagenomics analyses of the fecal samples from the same groups highlighted that athletes had a different microbiome composition also from a functional point of view, with increased microbial representation of genes involved in carbohydrate and amino acid metabolism, and short-chain fatty acid (SCFA) production (7). In another study, the average abundance of taxa involved in energy and carbohydrate metabolism, including *Prevotella* and *Methanobrevibacter smithii*, resulted significantly higher in professional than amateur cyclists, and was correlated with the frequency of training (89). However, these studies could not fully disentangle the contribution of exercise and diet in determining different microbiota compositions in different groups, since participants followed a wide range of dietary regimens. The intensity of training is also important: light exercise programs induce only subtle modifications of gut microbiota composition in sedentary subjects (31). Therefore, the findings of studies performed in athletes should not automatically be transferred to all subjects undertaking non-competitive exercise.

According to three different studies (37,38,127), fecal microbiota biodiversity is correlated with cardiorespiratory fitness in adult subjects. However, in one of these studies, performed in 71 premenopausal Finnish women, this relationship was mediated by body composition (127). Another study, performed in 19 active and 21 sedentary women aged ≤40 years old, confirmed that the microbiome abundance of several bacterial taxa was significantly correlated with the body fat or lean mass percentage (15). Thus, the possible association between exercise and microbiota should be further investigated, carefully taking into account possible confounders, such as dietary habits, nutrient intake, and parameters of body composition.

The influence of body composition on microbiota was emphasized also by the findings of one intervention study, where two groups of sedentary subjects, one lean and one obese, underwent a 6-week structured exercise program, followed by a 6-week washout period (2). After exercise training, both lean and obese participants experienced a change in gut microbiota composition, but the overall representation of species with known anti-inflammatory properties and the microbiome capacity of producing SCFA was higher in lean subjects, highlighting a body mass index (BMI)-dependent response to training. However, all the changes reversed towards the baseline status after the washout period (2). Interestingly, in exercised healthy young males undergoing a period of forced inactivity, cessation of exercise was associated with changes in gastrointestinal physiology (i.e. reduction of bowel movements and increased consistency of feces) before alterations of gut microbiota composition and function could be detected (103,104,105). These circumstances suggest that the microbiome is resilient to acute changes in exercise habits, and that maintenance of exercise is needed to induce long-lasting modifications of intestinal microbial ecosystem.

The modifications of intestinal microbiome composition induced by exercise can exert beneficial effects on the whole organism, modulating pathological processes. For example, exercise-induced microbiota changes are able to attenuate the

clinical course and outcome of experimental models of myocardial infarction or chemically-induced colitis, especially by modulating the inflammatory response (1,63). The key mediators in these processes may be SCFA, and particularly butyrate, whose production by gut microbiota has been shown to increase after exercise in humans (2).

2.2 Negative effects of exercise on gut microbiota

Despite the findings of the studies discussed above, other investigations have questioned the concept that the exercise-induced changes in gut microbiota composition are always favorable for the host physiology. Endurance high-intensity exercise, especially if not proportioned to training level, may in fact represent a huge stressor for the organism. These conditions can induce ischemic events in the gut mucosa, associated with acute gastrointestinal symptoms including abdominal pain, nausea, and diarrhea (32). From a gut microbiota perspective, these phenomena may be associated with increased intestinal permeability allowing several bacteria and their toxic products to enter systemic circulation and activate systemic inflammation (51,97). A basic mediator in these processes is represented by microbiota-derived lipopolysaccharide (LPS) (97), exerting a wide range of pathological actions on the host (4).

Moreover, the high-intensity exercise-induced dysfunction of the intestinal mucosa may promote profound and rapid changes in microbiota. For example, in a group of soldiers, a 4-day military training program of Arctic cross-country ski-march resulted in deep changes in fecal microbiota composition and functionality. Namely, there was an expansion of a large number of taxa, including opportunistic pathogens, at the expense of dominant taxa, such as *Bacteroides*, and taxa with known production of anti-inflammatory mediators (51). In amateur athletes, the fecal microbiome functionality acutely changed after a half-marathon race, exhibiting a pro-inflammatory profile with a completely different fecal metabolome (131). Similar changes have also been demonstrated in animal models (129). Interestingly, the administration of probiotics or prebiotics seems to attenuate these unfavorable changes of gut microbiota after exercise to exhaustion (22,48,97), although the benefits are uncertain in case of lower intensity of exercise (118,119).

In summary, regular exercise training seems to be associated with higher biodiversity and beneficial functions of intestinal microbiome. The microbiota may thus represent a mediator of the exercise-induced health benefits, although diet and body composition may play a relevant role in this association. On the other side, there are also some studies supporting that exercise to exhaustion may be associated with detrimental consequences for the microbio-

me. The effects of exercise on the intestinal microbiome may thus depend on its intensity and timing, and future studies should help to disentangle this relationship.

3. Can the microbiota influence muscle pathophysiology?

The gut-muscle axis hypothesis in age-related sarcopenia

Recently, several research groups have independently hypothesized that the gut microbiota composition may influence the emergence of sarcopenia, i.e. the loss of muscle mass and function occurring with aging (33,44,83,90,112,116). A study performed on rat models of sarcopenia has actually demonstrated that age-related muscle mass wasting is associated with a distinct fecal microbiota composition, with reduced representation of several taxa with purported anti-inflammatory and pro-anabolic actions on the host tissues, including *Clostridium XIVa* cluster, *Butyrivococcus*, *Sutterella*, *Coprococcus* and *Faecalibacterium* (102). Sarcopenic rats also exhibited a different fecal microbiota functionality, with rearrangements in the expression of bacterial genes involved in nutrient biosynthesis and catabolism (102).

To date, no studies have investigated the composition and functionality of fecal microbiota in older humans with sar-

Table 1: Overview of the possible pathophysiological mechanisms involved in the gut microbiota-mediated regulation of skeletal muscle function (gut-muscle axis) and of the corresponding effects in case of gut microbiota dysbiosis.

Hypothesized pathways involved in gut microbiota modulation of muscle function	Possible skeletal muscle effects in case of dysbiosis
Bioavailability of dietary proteins and specific amino acids (tryptophan) (36,62,101)	↓ protein synthesis ↓ IGF-1-mediated anabolic effect
Synthesis of vitamins, including folate, vitamin B12, riboflavin (59,111)	↓ DNA synthesis, methylation and repair ↓ oxidative stress neutralization capacity ↓ amino acid biosynthesis ↓ energy production
Biotransformation of nutrients, including polyphenols (resveratrol) and ellagitannins (97,106)	↓ mitochondrial biogenesis ↓ exercise resistance capacity ↓ oxidative stress neutralization capacity
Permeability of intestinal mucosa (20,25)	↑ inflammation (stimulation of TLRs) ↑ muscle protein catabolism ↓ insulin sensitivity with anabolic resistance
Biotransformation of bile acids (53)	↓ activation of anabolic pathways regulated by farnesoid X receptor
Synthesis of short-chain fatty-acids (19,118,123)	↓ stimulation of anabolism ↓ insulin sensitivity ↑ systemic and local pro-inflammatory cytokine production

IGF-1 = Insulin-Like Growth-Factor-1; TLR = Toll-Like Receptors

copenia (116). However, there is some indirect evidence supporting the hypothesis of a gut-muscle axis, in which the intestinal microbiota composition can influence muscle mass anabolism and functionality.

The physio-pathological substrates of sarcopenia are represented by reduced muscle capillarity, reduced insulin sensitivity, and increased subclinical inflammation, resulting in altered mitochondrial biogenesis and function, and altered anabolic/catabolic balance of muscle protein synthesis (14,69,70). From a clinical point of view, muscle mass loss may also be favored by several conditions that are frequently found in geriatric patients, including malnutrition, low dietary protein intake, intestinal malabsorption, altered digestion and subclinical cognitive deficits (58,83).

In this context, the intestinal microbiota composition may influence the onset of sarcopenia at multiple levels. The presence of gut microbiota dysbiosis is in fact associated with several metabolic alterations, involving protein synthesis, release of pro-anabolic mediators, inflammation and insulin sensitivity. All these elements can modulate skeletal muscle physiology, as summarized in Table 1.

First, a dysbiotic intestinal microbiota can reduce the bioavailability of dietary proteins (102) and particularly of some amino acids, like tryptophan, involved in modulation of inflammation and promotion of muscle protein synthesis (21,36,62). Gut bacteria are also involved in the synthesis of many vitamins, including folate, vitamin B12 and riboflavin, exerting several beneficial and pro-anabolic effects in skeletal muscle cells, ranging from amino acid biosynthesis to oxidative stress neutralization during exercise (59).

Moreover, a healthy intestinal microbiota can effectively transform some dietary nutrients into metabolic mediators that, once absorbed into systemic circulation, can exert beneficial effects on inflammation, insulin sensitivity, anabolism, and antioxidant capacity. Conversely, a dysbiotic microbiota may lack these functions, with some negative consequences on muscle health. Polyphenols, including resveratrol, and ellagitannins contained in pomegranates and berries represent the most relevant examples of nutrients that, after microbial metabolism, enter systemic circulation and exert beneficial effect for the muscle (98,107). Interestingly, endurance training seems to enhance the bioavailability of dietary polyphenols, probably through its beneficial modulations of intestinal microbiota (88).

Moreover, the age-related alterations of gut microbiota composition (24), occurring independently from the level of exercise training, can promote gut mucosa dysfunction, with increased permeability. This phenomenon may result in the systemic absorption of microbial byproducts and toxins, including LPS (20). In skeletal muscle cells, circulating LPS can contribute to activate Toll-Like Receptors (TLR) 4 and 5, promoting NF- κ B pathway activation, with reduced insulin sensitivity, enhanced protein catabolism and inflammatory cytokine production (72,106). In animal models, TLR4 activation determines muscle atrophy (35). In aging human beings, TLR4 activation is associated with metabolic endotoxemia, decreased insulin sensitivity and reduced quadriceps muscle strength and volume (41).

But probably the most studied mechanism involved in gut microbiota modulation of muscle function is the bacterial production of metabolic mediators, including bile acids and

SCFA (20,25). A healthy gut microbiota can produce secondary bile acids, that are well known activators of farnesoid X receptor stimulating myocyte anabolism (53). SCFA, and particularly butyrate, are generally synthesized by a large number of gut bacteria, including *Faecalibacterium*, *Butyricimonas*, and *Succinivibrio*, highly represented in healthy subjects but with reduced abundance in older individuals (19). These mediators have several beneficial metabolic activities, summarized in Table 2, ultimately influencing skeletal muscle protein deposition through modulation of the systemic anabolic/catabolic balance (6,19). The administration of butyrate and probiotics with similar functionality to animal models of muscle wasting resulted in massive improvements in muscle mass (120,125). Unfortunately, human studies on this topic are still lacking to date.

The metabolic action of gut microbiota was however confirmed in a study by Blanton and colleagues, where the transplantation of the dysbiotic fecal microbiota from malnourished African children to germ-free mice resulted in mouse failure-to-thrive (12).

Furthermore, the administration of rifaximin to mouse models of hepatic encephalopathy surprisingly resulted in improved skeletal muscle mass and function (56). Rifaximin is able to

Table 2: Summary of the main physiological functions of short-chain fatty acids (butyrate, acetate, propionate) produced by the intestinal microbiota (6,19,118,123). The most relevant functions possibly involved in the gut-muscle axis are shown in italics.

Substance	Function
Acetate	<i>Direct modulation of systemic inflammation</i>
	<i>Antagonization of LPS-driven inflammation</i>
	<i>Improvement in insulin sensitivity</i>
	<i>Stimulation of skeletal muscle glucose uptake</i>
	Peripheral modulation of satiety via GLP-1 and PYY
	Central appetite modulation
	Promotion of lipolysis
	Induction of adipose tissue differentiation
	Increased energy expenditure via thermogenesis
Propionate	<i>Increased Treg cell differentiation</i>
	<i>Direct modulation of systemic inflammation</i>
	<i>Antagonization of LPS-driven inflammation</i>
	Peripheral modulation of satiety via GLP-1 and PYY
	Promotion of lipolysis
	Induction of adipose tissue differentiation
Butyrate	<i>Direct modulation of systemic inflammation</i>
	<i>Antagonization of LPS-driven inflammation</i>
	<i>Improvement in insulin sensitivity</i>
	<i>Stimulation of skeletal muscle glucose uptake</i>
	<i>Histone deacetylase inhibition</i>
	<i>Reduced intestinal permeability</i>
	Peripheral modulation of satiety via GLP-1 and PYY
	Central appetite modulation
	Promotion of lipolysis
	Induction of adipose tissue differentiation
Increased energy expenditure via thermogenesis	

GLP-1 = Glucagon-Like Peptide-1; PYY = Peptide YY; LPS = Lipopolysaccharide

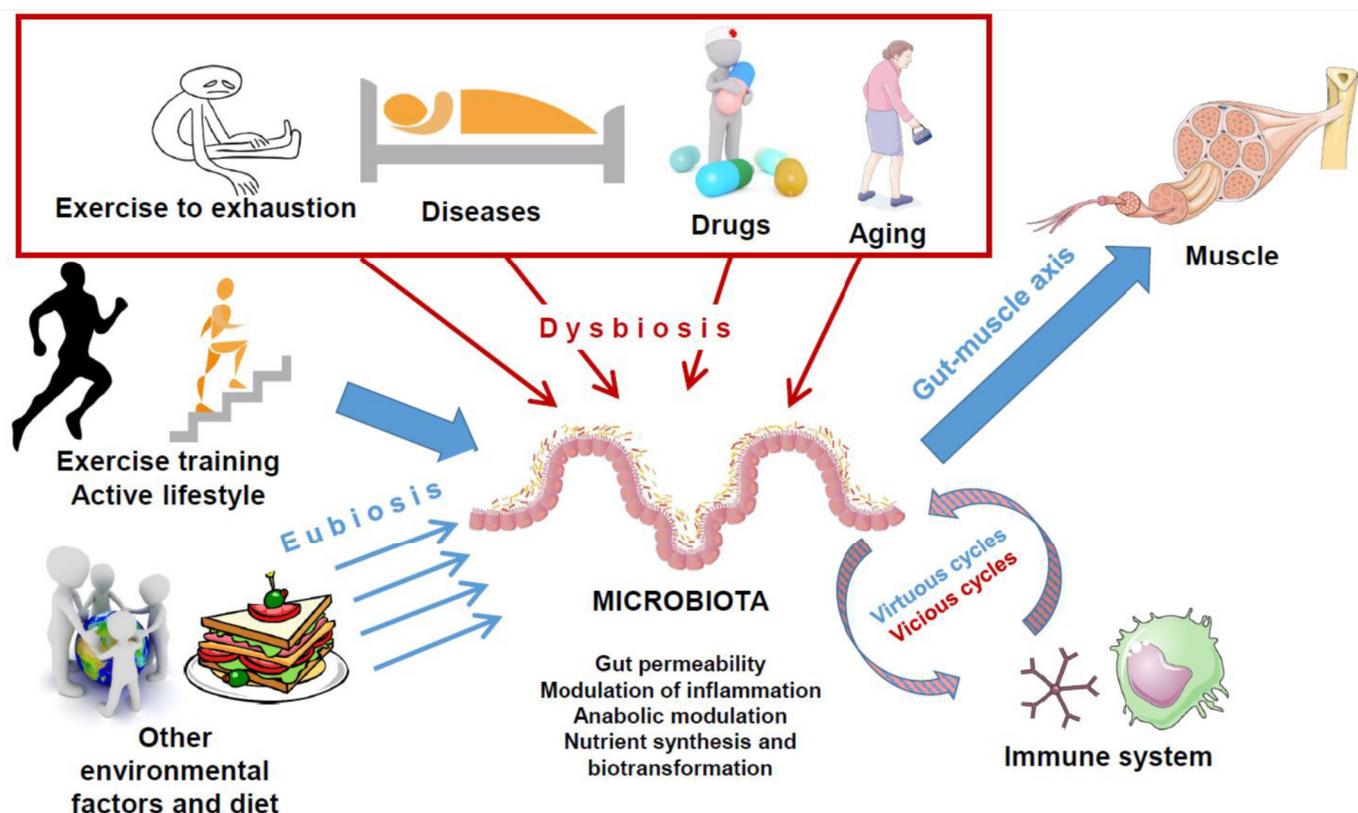


Figure 1: Representation of the hypothetical conceptual framework connecting exercise, intestinal microbiome, inflammation, immune system function and skeletal muscle pathophysiology at the current literature state of art.

selectively kill pathobionts, favoring the expansion of bacterial populations with purported beneficial activities, such as *Bifidobacteria* or *Lactobacilli* (91). The observed effects on mice may depend on its capacity to reduce the gut microbiota dysbiosis associated with hepatic encephalopathy.

In summary, the results of several pre-clinical studies support the hypothesis that gut microbiota dysbiosis may be associated with muscle wasting, especially in age-related sarcopenia. However, confirmation of this possible gut-muscle axis in human studies is still lacking, and the clinical relevance of these supposed mechanisms is still uncertain.

4. Exercise and the gut-muscle axis

Although studies on humans are lacking, several preclinical studies support the hypothesis that the intestinal microbiota can modulate skeletal muscle physiology not only in age-related sarcopenia, but in all ages and physiological states (20,25). In this context, exercise may represent a strong modulator of gut microbiota composition. Thus, the gut-muscle communication in human pathophysiology may be bidirectional (25), with gut microbiota representing a “cross-road” among environment, lifestyle, and skeletal muscle (112).

In this scenario, some authors have hypothesized that many of the well-known positive health effects of exercise may be mediated by its beneficial modifications on the gut microbiota (23,79,80). However, when there is an exercise overload, these possible beneficial effects are outweighed by increased intestinal permeability and oxidative stress, promoting inflammation and a catabolic state that negatively impacts the functionality of skeletal muscle (29). Moreover, the harmful

effects of inactivity on the muscle and vascular system may be at least partly mediated by negative changes of the gut microbiota towards dysbiosis (23,109).

In healthy subjects who regularly perform physical activity, a homeostatic equilibrium between intestinal microbiota and skeletal muscle may be present, with exercise promoting healthy microbiota composition, and microbiota favoring muscle health. This equilibrium may be disrupted by sedentary lifestyle or excessive exercise, resulting in dysbiosis of the gut microbiota. Other factors promoting dysbiosis, such as drugs or acute illnesses, may also be associated with reduced muscle mass and function. In fact, dysbiosis influences gut permeability, systemic inflammation, anabolism and nutrient availability. All these mechanisms are involved in muscle physiology and represent the substrates of the gut-muscle axis, as depicted in Figure 1.

In summary, the gut-muscle axis may be two-way, with microbiota influencing the muscle, and exercise contributing to shape microbiota composition. The intensity and frequency of exercise may have great importance in determining which way of the axis is prevalent, and its physio-pathological consequences.

5. Immune system and the gut-muscle axis: virtuous and vicious cycles

A healthy gut microbiota has a fundamental role in shaping local and systemic immune response to gut bacteria through the whole lifespan, favoring the maintenance of tolerance towards antigens from commensals and activation against antigens from pathogens (40). On the other side, gut micro-

biota dysbiosis favors the loss of immunologic tolerance to commensals, the impairment of epithelial barrier function and an imbalance in the activation of anti-inflammatory T_{reg} lymphocytes and pro-inflammatory Th17 lymphocytes (43,60). These phenomena may contribute to the onset of several infectious, inflammatory and autoimmune diseases, including inflammatory bowel diseases, type 1 diabetes and multiple sclerosis, with gut microbiota playing an active pathogenic role (9).

However, besides this “outside-in” relationship, there is also an important “inside-out” control of immune system over gut microbiota (47,117). The immune system is in fact able to influence the gut microbiota composition at multiple levels. Both innate and adaptive immunity are involved. The possible mechanisms are synthesized in Table 3, and include production of antimicrobial peptides from intestinal cells, mucus secretion, immunoglobulin A (IgA) activation, toll-like receptor (TLR) activation, lymphocyte transfer and differentiation, presence of invariant natural killer T cells (iNKT) (47,60,68,101,130). The presence of specific imbalances in each one of these pathways may be associated with the emergence of gut microbiota dysbiosis (47,68).

In fact, human beings infected with the Human Immunodeficiency Virus (HIV) exhibit deep changes in the structure and functionality of the intestinal microbiome, with increased biodiversity due to overgrowth of opportunistic pathogens and decreased representation of taxa with anti-inflammatory properties (121,122,133). Similar alterations of gut microbial community structure have been detected also in patients with IgA deficiency (39). These findings support the concept that the immune system functionality influences gut microbiota composition.

Conversely, the presence of specific functionalities in the gut microbiome, related to fatty acid metabolism, PPAR-signaling, lipid biosynthesis and kynurenine pathway of tryptophan metabolism, may enhance systemic immunity activation and promote control of HIV infection (124). Therefore, a complex interplay between microbiome and immunity exists, and the physio-pathological consequences depend on the type of equilibrium reached (117).

Some metabolic mediators, such as bile salts, may play a relevant role in this equilibrium. In the gut lumen, bile salts can in fact undergo metabolic transformations into compounds with immunoregulatory and anti-inflammatory properties, particularly on Kupffer cells and intrahepatic lymphocytes (100). Bile salts have also the capacity of selecting specific subpopulations of the gut microbiota that are able to metabolize them, contributing to shape the intestinal microenvironment (100).

Table 3: Overview of the mechanisms involved in immune system control of gut microbiota composition (47,60,68,100,128).

Immunity type	Mechanism
Intestinal innate immunity	Production of mucus (barrier function)
	Production of antimicrobial peptides by Paneth cells
	Production of α -defensins by epithelial cells
	Activation of NOD-like receptor and production of IL-18 by epithelial cells
	Release of non-specific immunoglobulins
	Invariant Natural Killer lymphocyte activation
	Expression of Resistin-like molecule β
Leptin expression	
Intestinal acquired immunity	IgA response to gut microbiota antigens
	Activation of CD4+ T cells in intestinal mucosa
	Activation of Foxp3+ T cells in intestinal mucosa
	Activation of Toll-Like Receptors with lymphocyte stimulation
HLA class I and II loci expression	
Systemic acquired immunity	IgG response to gut microbiota antigens penetrated in systemic circulation

Table 4: Summary of the main features of aging immune system involved in increased gut mucosa permeability and in age-related gut microbiota dysbiosis (11,66,124).

Immunity type	Age-related alterations
Innate	Reduced Paneth cell function
	Reduced glycosylation of mucins
	Reduction of M cell number and function
	Increased levels of pro-inflammatory cytokines IL-1 β , IL-6 and TNF- α
	Decreased levels of anti-inflammatory cytokines including IL-10
	Increased production of adipokines
	Impaired phagocytic function of macrophages
	Overgrowth of pro-inflammatory CX3CR1 ^{int} macrophages
	Reduced chemotactic capacity in neutrophils
	Reduced antigen presentation capacity by dendritic cells
Acquired	Reduced T cell priming by dendritic cells
	Reduced differentiation of CD4+ CD25+ T cells
	Reduced T cell secretion of TGF- β
	Increased T cell secretion of IFN- γ
	Overgrowth of T _{reg} lymphocytes
	Reduced homing of IgA-secreting B lymphocytes
	Reduced IgA secretion capacity
	Loss of naïve T and B cells

Whatever the mediators involved, the equilibrium between immune system and microbiome may be strongly influenced by environmental factors. Positive modulators of gut microbiota composition, including regular exercise, may induce a beneficial equilibrium with the immune system, resulting in a virtuous cycle helping to maintain health (27,85). Conversely, factors that disrupt gut microbiota composition, such as exercise to exhaustion, illness and aging, may cause a perturbation of the equilibrium between microbiome and immune system. As a result, systemic inflammation is chronically activated, sustaining further alterations of the microbiota towards dysbiosis promoted by the altered immune system regulation (30,46). So, a vicious circle arises.

These postulated mechanisms are highlighted in Figure 1.

A healthy gut microbiota, and a positive interaction with the immune system, may be crucial for the gut-muscle axis, and may influence the maintenance of muscle mass and functionality, especially in exercised subjects (29). Conversely, dysbiosis resulting from a negative interaction with the immune system may influence muscle wasting disorders, particularly during aging (112).

Age-related gut microbiota dysbiosis is associated with increased gut mucosa permeability in both animal models and humans (92,111). The reduced intestinal epithelial barrier function is accompanied by several alterations in immune system, involving both innate and acquired immunity (Table 4) (11,66,126). These alterations ultimately promote local and systemic inflammation, with overproduction of the pro-inflammatory cytokines TNF- α , IL-1 β and IL-6 (66,126). Inflammation negatively impacts the gut-muscle axis and is involved in the pathogenesis of several age-related conditions, including not only sarcopenia and frailty (126), but even cancer (11). Moreover, IL-1 β further stimulates intestinal epithelial tight junction permeability and promotes local dysbiosis (3), in a vicious cycle supporting skeletal muscle wasting and loss of function.

In summary, the relationship between intestinal microbiome and immunity may be two-way, and the resulting equilibrium may exert important functions on the functionality of the gut-muscle axis and on muscle health. More research is however needed to disentangle these complex relationships, and to reveal their actual relevance from a clinical perspective.

6. Conclusions

The relationship between exercise, immune system, gut microbiota, and skeletal muscle pathophysiology is very complex and not completely elucidated at the current state of the art. In Figure 1, we present a possible interpretative framework, showing that the gut microbiota is at the cross-road between environmental stimuli and host physiology, undergoing a continuous interplay with the immune system and the skeletal muscle.

Future studies should clarify whether gut microbiota dysbiosis is pathophysiologically associated with muscle wasting disorders, and if exercise may positively influence this putative gut-muscle axis. Furthermore, the influence of the microbiome-immune system interplay on skeletal muscle mass and functionality should be investigated in both experimental models and human beings.

ACKNOWLEDGMENTS

Part of the images are distributed under Creative Commons Licence and can be freely available at the following links: <https://smart.servier.com/> and <https://pixabay.com>.

The authors wish to thank Dr. Cristoforo Incorvaia for advice on manuscript writing.

FINANCIAL SUPPORT AND SPONSORSHIP

None.

COMPETING INTERESTS

All the authors have no conflict of interest to declare.

REFERENCES

- Allen JM, Mailing LJ, Cohrs J, Salmonson C, Fryer JD, Nehra V, Hale VL, Kashyap P, White BA, Woods JA. Exercise training-induced modification of the gut microbiota persists after microbiota colonization and attenuates the response to chemically-induced colitis in gnotobiotic mice. *Gut Microbes* 9: 115-130, 2018.
- Allen JM, Mailing LJ, Niemi GM, Moore R, Cook MD, White BA, Holscher HD, Woods JA. Exercise alters gut microbiota composition and function in lean and obese humans. *Med Sci Sports Exerc* 50: 747-757, 2018.
- Al-Sadi R, Ye D, Said HM, Ma TY. IL-1 β -induced increase in intestinal epithelial tight junction permeability is mediated by MEKK-1 activation of canonical NF- κ B pathway. *Am J Pathol* 177: 2310-2322, 2010.
- Armstrong LE, Lee EC, Armstrong EM. Interactions of gut microbiota, endotoxemia, immune function, and diet in exertional heatstroke. *J Sports Med* 2018: 5724575, 2018.
- Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, Fernandes GR, Tap J, Bruls T, Batto JM, Bertalan M, Borruel N, Casellas F, Fernandez L, Gautier L, Hansen T, Hattori M, Hayashi T, Kleerebezem M, Kurokawa K, Leclerc M, Levenez F, Manichanh C, Nielsen HB, Nielsen T, Pons N, Poulain J, Qin J, Sicheritz-Ponten T, Tims S, Torrents D, Ugarte E, Zoetendal EG, Wang J, Guarner F, Pedersen O, de Vos WM, Vrunak S, Doré J, MetaHIT Consortium. Enterotypes of the human gut microbiome. *Nature* 473: 174-180, 2011.
- Backhed F, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci USA* 104: 979-984, 2007.
- Barton W, Penney NC, Cronin O, Garcia-Perez I, Molloy MG, Holmes E, Shanahan F, Cotter PD, O'Sullivan O. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut* 67: 625-633, 2018.
- Batacan RB, Fenning AS, Dalbo VJ, Scanlan AT, Duncan MJ, Moore RJ, Stanley D. A gut reaction: the combined influence of exercise and diet on gastrointestinal microbiota in rats. *J Appl Microbiol* 122: 1627-1638, 2017.
- Belkaid Y, Hand TW. Role of microbiota in immunity and inflammation. *Cell* 157: 121-141, 2014.
- Bermon S, Petriz B, Kajeniene A, Prestes J, Castell L, Franco OL. The microbiota: an exercise immunology perspective. *Exerc Immunol Rev* 21: 70-79, 2015.
- Biragyn A, Ferrucci L. Gut dysbiosis: a potential link between increased cancer risk in ageing and inflammaging. *Lancet Oncol* 19: e295-e304, 2018.
- Blanton LV, Charbonneau MR, Salih T, Barratt MJ, Venkatesh S, Ilkaveya O, Subramanian S, Manary MJ, Trehan I, Jorgensen JM, Fan YM, Henrissat B, Leyn SA, Rodionov DA, Osterman AL, Maleta KM, Newgard CB, Ashorn P, Dewey KG, Gordon JI. Gut bacteria that prevent growth impairments transmitted by microbiota from malnourished children. *Science* 351: aad3311, 2016.

13. Bonder MJ, Kurilshikov A, Tigchelaar EF, Mujagic Z, Imhann F, Vila AV, Deelen P, Vatanen T, Schirmer M, Smeekens SP, Zhernakova DV, Jankipersadsing SA, Jaeger M, Oosting M, Cenit MC, Masclee AA, Swertz MA, Li Y, Kumar V, Joosten L, Harmsen H, Weersma RK, Franke L, Hofker MH, Xavier RJ, Jonkers D, Netea MG, Wijmenga C, Fu J, Zhernakova A. The effect of host genetics on the gut microbiome. *Nat Genet* 48: 1407-1412, 2016.
14. Bowen TS, Schuler G, Adams V. Skeletal muscle wasting in cachexia and sarcopenia: molecular pathophysiology and impact of exercise training. *J Cachexia Sarcopenia Muscle* 6: 197-207, 2015.
15. Bressa C, Bailen-Andrino M, Perez-Santiago J, Gonzalez-Soltero R, Perez M, Montalvo-Lominchar MG, Maté-Munoz JL, Dominguez R, Moreno D, Larrosa M. Differences in gut microbiota profile between women with active lifestyle and sedentary women. *PLoS One* 12: e0171352, 2017.
16. Buttò LF, Schaebeck M, Haller D. Mechanisms of microbe-host interaction in Crohn's disease: dysbiosis vs pathobiont selection. *Front Immunol* 6: 555, 2015.
17. Calvani R, Picca A, Lo Monaco MR, Landi F, Bernabei R, Marzetti E. Of microbes and minds: an narrative review on the second brain aging. *Front Med* 5: 53, 2018.
18. Campbell SC, Wisniewski PJ, Noji M, McGuinness LR, Haggblom MM, Lightfoot SA, Joseph LB, Kerkhof LJ. The effect of diet and exercise on intestinal integrity and microbial diversity in mice. *PLoS One* 11: e0150502, 2016.
19. Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol* 11: 577-591, 2015.
20. Cerdà B, Perez M, Perez-Santiago JD, Tornero-Aguilera JF, Gonzalez-Soltero R, Larrosa M. Gut microbiota modification: another piece in the puzzle of the benefits of physical exercise in health? *Front Physiol* 7: 51, 2016.
21. Cervenka I, Agudelo LZ, Ruas JL. Kynurenines: tryptophan's metabolites in exercise, inflammation, and mental health. *Science* 357: 369, 2017.
22. Chaves FM, Baptista IL, Simabuco FM, Quaresma PGF, Pena FL, Bezerra RMN, Pauli JR, da Cunha DT, Campos-Ferraz PL, Antunes AEC. High-intensity-exercise-induced intestinal damage is protected by fermented milk supplemented with whey protein, probiotic and pomegranate (*Punica granatum* L.). *Br J Nutr* 119: 896-909, 2018.
23. Chen J, Guo Y, Gui Y, Xu D. Physical exercise, gut, gut microbiota, and atherosclerotic cardiovascular diseases. *Lipids Health Dis* 17: 17, 2018.
24. Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, Harris HM, Coakley M, Lakshminarayanan B, O'Sullivan O, Fitzgerald GF, Deane J, O'Connor M, Harnedy N, O'Connor K, O'Mahony D, van Sinderen D, Wallace M, Brennan L, Stanton C, Marchesi JR, Fitzgerald AP, Shanahan F, Hill C, Ross RP, O'Toole PW. Gut microbiota composition correlates with diet and health in the elderly. *Nature* 488: 178-184, 2012.
25. Clark A, Mach N. The crosstalk between the gut microbiota and mitochondria during exercise. *Front Physiol* 8: 319, 2017.
26. Clarke SF, Murphy EF, O'Sullivan O, Lucey AJ, Humphreys M, Hogan A, Hayes P, O'Reilly M, Jeffery IB, Wood-Martin R, Kerins DM, Quigley E, Ross RP, O'Toole PW, Molloy MG, Falvey E, Shanahan F, Cotter PD. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut* 63: 1913-1920, 2014.
27. Codella R, Luzi L, Terruzzi I. Exercise has the guts: how physical activity may positively modulate gut microbiota in chronic and immune-based diseases. *Dig Liver Dis* 50: 331-341, 2018.
28. Conlon MA, Bird AR. The impact of diet and lifestyle on gut microbiota and human health. *Nutrients* 7: 17-44, 2014.
29. Cook MD, Allen JM, Pence BD, Wallig MA, Gaskins HR, White BA, Woods JA. Exercise and gut immune function: evidence of alterations in colon immune cell homeostasis and microbiome characteristics with exercise training. *Immunol Cell Biol* 94: 158-163, 2016.
30. Cook MD, Martin SA, Williams C, Whitlock K, Wallig MA, Pence BD, Woods JA. Forced treadmill exercise training exacerbates inflammation and causes mortality while voluntary wheel training is protective in a mouse model of colitis. *Brain Behav Immun* 33: 46-56, 2013.
31. Cronin O, Barton W, Skuse P, Penney NC, Garcia-Perez I, Murphy EF, Woods T, Nugent H, Fanning A, Melgar S, Falvey EC, Holmes E, Cotter PD, O'Sullivan O, Molloy MG, Shanahan F. A prospective metagenomics and metabolomics analysis of the impact of exercise and/or whey protein supplementation on the gut microbiome of sedentary adults. *mSystems* 3: e00044-18, 2018.
32. de Oliveira EP, Burini R. The impact of physical exercise on the gastrointestinal tract. *Curr Opin Clin Nutr Metab Care* 12: 533-538, 2009.
33. de Sire R, Rizzatti G, Ingravalle F, Pizzoferrato M, Petito V, Lopetuso L, Graziani C, de Sire A, Mentella MC, Mele MC, Gasbarrini A, Scaldaferrri F. Skeletal muscle-gut axis: emerging mechanisms of sarcopenia for intestinal and extra-intestinal diseases. *Minerva Gastroenterol Dietol* 64: 351-362, 2018.
34. Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, Knight R. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA* 107: 11971-11975, 2010.
35. Doyle A, Zhang G, Abdel Fattah EA, Eissa NT, Li YP. Toll-like receptor 4 mediates lipopolysaccharide-induced muscle catabolism via coordinate activation of ubiquitin-proteasome and autophagy-lysosome pathways. *FASEB J* 25: 99-110, 2011.
36. Dukes A, Davis C, El Refaey M, Upadhyay S, Mork S, Arounleut P, Johnson MH, Hill WD, Isales CM, Hamrick MW. The aromatic amino acid tryptophan stimulates skeletal muscle IGF1/p70s6k/mTor signaling in vivo and the expression of myogenic genes in vitro. *Nutrition* 31: 1018-1024, 2015.
37. Durk RP, Castillo E, Marquez-Megana L, Grosicki GJ, Bolter ND, Lee CM, Bagley JR. Gut microbiota composition is related to cardiorespiratory fitness in healthy young adults. *Int J Sport Nutr Exerc Metab* ahead of print Jun 10, 2018. Doi: 10.1123/ijsnem.2018-0024
38. Estaki M, Pither J, Baumeister P, Little JP, Gill SK, Ghosh S, Ahmadi-Vand Z, Marsden KR, Gibson DL. Cardiorespiratory fitness as a predictor of intestinal microbial diversity and distinct metagenomic functions. *Microbiome* 4: 42, 2016.
39. Fadlallah J, El Kafsi H, Sterlin D, Juste C, Parizot C, Dorgham K, Autaa G, Gouas D, Almeida M, Lepage P, Pons N, Le Chatelier E, Levenez F, Kennedy S, Galleron N, Pais de Barros JP, Malphettes M, Galicier L, Boutboul D, Mathian A, Miyara M, Oksenhendler E, Amoura Z, Doré J, Fieschi C, Ehrlich SD, Larsen M, Gorochov G. Microbial ecology perturbation in human IgA deficiency. *Sci Transl Med* 10: eaan1217, 2018.

40. Gensollen T, Iyer SS, Kasper DL, Blumberg RS. How colonization by microbiota in early life shapes the immune system. *Science* 352: 539-544, 2016.
41. Ghosh S, Lertwattanarak R, de Jesus Garduno J, Joya Galeana J, Li J, Zamarripa F, Lancaster JL, Mohan S, Hussey S, Musi N. Elevated TLR4 expression and metabolic endotoxemia in human aging. *J Gerontol A Biol Sci Med Sci* 70: 232-246, 2015.
42. Greenhalgh K, Meyer KM, Aagaard KM, Wilmes P. The human gut microbiome in health: establishment and resilience of microbiota over a lifetime. *Environ Microbiol* 18: 2103-2116, 2016.
43. Grigg JB, Sonnenberg GF. Host-microbiota interactions shape local and systemic inflammatory diseases. *J Immunol* 198: 564-571, 2017.
44. Grosicki GJ, Fielding RA, Lustgarten MS. Gut microbiota contribute to age-related changes in skeletal muscle size, composition, and function: biological basis for a gut-muscle axis. *Calcif Tissue Int* 102: 433-442, 2018.
45. Guarner F, Malagelada JR. Gut flora in health and disease. *Lancet* 361: 512-519, 2003.
46. Hofmann-Goetz H, Quadrilatero J. Treadmill exercise in mice increases intestinal lymphocyte loss via apoptosis. *Acta Physiol Scand* 179: 289-297, 2003.
47. Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. *Science* 336: 1268-1273, 2012.
48. Hsu YJ, Huang WC, Lin JS, Chen YM, Ho ST, Huang CC, Tung YT. Kefir supplementation modifies gut microbiota composition, reduces physical fatigue, and improves exercise performance in mice. *Nutrients* 10: 862, 2018.
49. Jackson MA, Jeffery IB, Beaumont M, Bell JT, Clark AG, Ley RE, O'Toole PW, Spector TD, Steves CJ. Signatures of early frailty in the gut microbiota. *Genome Med* 8: 8, 2016.
50. Kabat AM, Srinivasan N, Maloy KJ. Modulation of immune development and function by intestinal microbiota. *Trans Immunol* 35: 507-517, 2014.
51. Karl JP, Margolis LM, Madslie EH, Murphy NE, Castellani JW, Gundersen Y, Hoke AV, Levangie MW, Kumar R, Chakraborty N, Gautam A, Hammamieh R, Martini S, Mountain SJ, Pasiakos SM. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress. *Am J Physiol Gastrointest Liver Physiol* 312: G559-G571, 2017.
52. Karu N, Deng L, Slae M, Guo AC, Sajed T, Huynh H, Wine E, Wishart DS. A review on human fecal metabolomics: methods, applications, and the human fecal metabolome database. *Anal Chim Acta* 1030: 1-24, 2018.
53. Kobayashi Y, Hara N, Sugimoto R, Mifuji-Moroka R, Tanaka H, Eguchi A, Iwasa M, Hasegawa H, Iwata K, Takei Y, Taguchi O. The associations between circulating bile acids and the muscle volume in patients with non-alcoholic fatty liver disease (NAFLD). *Intern Med* 56: 755-762, 2017.
54. Koenig JE, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, Angenent LT, Rey RE. Succession of microbial consortia in the developing infant gut microbiome. *Proc Natl Acad Sci USA* 108S1: 4578-4585, 2011.
55. Kriss M, Hazleton KZ, Nusbacher NM, Martin CG, Lozupone CA. Low diversity gut microbiota dysbiosis: drivers, functional implications and recovery. *Curr Opin Microbiol* 44: 34-40, 2018.
56. Kumar A, Davuluri G, Nascimento e Silva R, Engelen MPKJ, Ten Have GAM, Prayson R, Deutz NEP, Darasathy S. Ammonia lowering reverses sarcopenia of cirrhosis by restoring skeletal muscle proteostasis. *Hepatology* 65: 2045-2058, 2017.
57. Lamoreux EV, Grandy SA, Langille MGI. Moderate exercise has limited but distinguishable effects on the mouse microbiome. *mSystems* 2: e00006-17, 2017.
58. Lauretani F, Maggio M, Ticinesi A, Tana C, Prati B, Gionti L, Nouvenne A, Meschi T. Muscle weakness, cognitive impairment and their interaction on altered balance in elderly outpatients: results from the TRIP observational study. *Clin Interv Aging* 13: 1437-1443, 2018.
59. LeBlanc JG, Milani C, de Giori GS, Sesma F, van Sinderen D, Ventura M. Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol* 24: 160-168, 2013.
60. Levy M, Kolodziejczyk AA, Thaïss CA, Elinav E. Dysbiosis and the immune system. *Nat Rev Immunol* 17: 219-232, 2017.
61. Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. *Cell* 124: 837-848, 2006.
62. Lin R, Liu W, Piao M, Zhu H. A review of the relationship between the gut microbiota and amino acid metabolism. *Amino Acids* 49: 2083-2090, 2017.
63. Liu Z, Liu HY, Zhou H, Zhan Q, Lai W, Zeng Q, Ren H, Xu D. Moderate-intensity exercise affects gut microbiome composition and influences cardiac function in myocardial infarction mice. *Front Microbiol* 8: 1687, 2017.
64. Logan AC, Jacka FN, Prescott SL. Immune-microbiota interactions: dysbiosis as a global health issue. *Curr Allergy Asthma Rep* 16:13, 2016.
65. Lozupone CA, Stombaugh JI, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature* 489: 220-230, 2012.
66. Man AL, Gicheva N, Nicoletti C. The impact of ageing on the intestinal epithelial barrier and immune system. *Cell Immunol* 289: 112-118, 2014.
67. Marchesi JR, Adams DH, Fava F, Hermes GD, Hirschfield GM, Hold G, Quraishi MN, Kinross J, Smidt H, Tuohy KM, Thomas LV, Zoetendal EG, Hart A. The gut microbiota and host health: a new clinical frontier. *Gut* 65: 330-339, 2016.
68. Marietta E, Rishi A, Taneja V. Immunogenetic control of the intestinal microbiota. *Immunology* 145: 313-322, 2015.
69. Martone AM, Marzetti E, Calvani R, Picca A, Tosato M, Santoro L, Di Giorgio A, Nesci A, Sisto A, Santoliquido A, Landi F. Exercise and protein intake: a synergistic approach against sarcopenia. *Biomed Res Int* 2017: 2672435, 2017.
70. Marzetti E, Calvani R, Tosato M, Cesari M, Di Bari M, Cherubini A, Collamati A, D'Angelo E, Pahor M, Bernabei R, Landi F, SPRINTT Consortium. Sarcopenia: an overview. *Aging Clin Exp Res* 29: 11-17, 2017.
71. McCoy KD, Ignacio A, Geuking MB. Microbiota and type 2 immune responses. *Curr Opin Immunol* 54: 20-27, 2018.
72. McFarlin BK, Flynn MG, Campbell WW, Stewart LK, Timmerman KL. TLR4 is lower in resistance-trained older women and related to inflammatory cytokines. *Med Sci Sports Exerc* 36: 1876-1883, 2004.
73. McKenzie C, Tan J, Macia L, Mackay CR. The nutrition-gut microbiome-physiology axis and allergic diseases. *Immunol Rev* 278: 277-295, 2017.

74. McLean MH, Dieguez jr D, Miller LM, Young HA. Does the microbiota play a role in the pathogenesis of autoimmune diseases? *Gut* 64: 332-341, 2015.
75. Mika A, Van Treuren W, Gonzalez A, Herrera JJ, Knight R, Fleshner M. Exercise is more effective at altering gut microbial composition and producing stable changes in lean mass in juvenile versus adult male F344 rats. *PLoS One* 10: e0125889, 2015.
76. Milani C, Ferrario C, Turrioni F, Duranti S, Mangifesta M, van Sinderen D, Ventura M. The human gut microbiota and its interactive connections to diet. *J Hum Nutr Diet* 29: 539-546, 2016.
77. Milani C, Hevia A, Foroni E, Duranti S, Turrioni F, Lugli GA, Sanchez B, Martin R, Gueimonde M, van Sinderen D, Margolles A, Ventura M. Assessing the fecal microbiota: an optimized ion torrent 16S rRNA gene-based analysis protocol. *PLoS One* 8: e68739, 2013.
78. Milani C, Mancabelli L, Lugli GA, Duranti S, Turrioni F, Ferrario C, Mangifesta M, Viappiani A, Ferretti P, Gorfer V, Tett A, Segata N, van Sinderen D, Ventura M. Exploring vertical transmission of Bifidobacteria from mother to child. *Appl Environ Microbiol* 81: 7078-7087, 2015.
79. Molina-Molina E, Lunardi Baccetto R, Wang DQH, de Bari O, Krawczyk M, Portincasa P. Exercising the hepatobiliary-gut axis. The impact of physical activity performance. *Eur J Clin Invest* 48: e12958, 2018.
80. Monda V, Villano I, Messina A, Valenzano A, Esposito T, Moscatelli F, Viggiano A, Cibelli G, Chieffi S, Monda M, Messina G. Exercise modifies the gut microbiota with positive health effects. *Oxid Med Cell Longev* 2017: 3831972, 2017.
81. Mosca A, Leclerc M, Hugot JP. Gut microbiota diversity and human diseases: should we reintroduce key predators in our ecosystem? *Front Microbiol* 7: 455, 2016.
82. Neish AS. Microbes in gastrointestinal health and disease. *Gastroenterology* 136: 65-80, 2009.
83. Ni Lochlainn M, Bowyer MCE, Steves CJ. Dietary protein and muscle in aging people: the potential role of gut microbiome. *Nutrients* 10: E929, 2018.
84. Ohlsson C, Sjogren K. Effects of the gut microbiota on bone mass. *Trends Microbiol* 26: 69-74, 2015.
85. O'Sullivan O, Cronin O, Clarke SF, Murphy EF, Molloy MG, Shanahan F, Cotter PD. Exercise and the microbiota. *Gut Microbes* 6: 131-136, 2015.
86. O'Toole PW, Jeffery IB. Gut microbiota and aging. *Science* 350: 1214-1215, 2015.
87. Pane M, Amoruso A, Deidda F, Graziano T, Allesina S, Mogna L. Gut microbiota, probiotics, and sport. From clinical evidence to agonistic performance. *J Clin Gastroenterol* 52: S46-S49, 2018.
88. Pereira-Cano G, Polyviou T, Ludwig IA, Nastase AM, Moreno-Rojas JM, Garcia AL, Malkova D, Crozier A. Bioavailability of orange juice (poly)phenols: the impact of short-term cessation of training by male endurance athletes. *Am J Clin Nutr* 106: 791-800, 2017.
89. Petersen LM, Bautista EJ, Nguyen H, Hanson BM, Chen L, Lek SH, Sodergren E, Weinstock GM. Community characteristics of the gut microbiomes of competitive cyclists. *Microbiome* 5: 98, 2017.
90. Picca A, Fanelli F, Calvani R, Mulè G, Pesce V, Sisto A, Pantanelli C, Bernabei R, Landi F, Marzetti E. Gut dysbiosis and muscle aging: searching for novel targets against sarcopenia. *Mediators Inflamm* 2018: 7026198, 2018.
91. Ponziani FR, Zocco MA, D'Aversa F, Pompili M, Gasbarrini A. Eubiotic properties of rifaximin: disruption of the traditional concepts in gut microbiota modulation. *World J Gastroenterol* 23: 4491-4499, 2017.
92. Qi Y, Goel R, Kim S, Richards EM, Carter CS, Pepine CJ, Raizada MK, Buford TW. Intestinal permeability biomarker zonulin is elevated in healthy aging. *J Am Med Dir Assoc* 18: 810.e1-810.e4, 2017.
93. Quince C, Walker AW, Simpson JT, Loman NJ, Segata N. Shotgun metagenomics, from sampling to analysis. *Nat Biotechnol* 35: 833-844, 2017.
94. Rajilic-Stojanovic M, de Vos WM. The first 1000 cultural species of the human intestinal microbiota. *FEMS Microbiol Rev* 36: 996-1047, 2014.
95. Ramezani A, Raj DS. The gut microbiome, kidney disease, and targeted interventions. *J Am Soc Nephrol* 25: 657-670, 2014.
96. Relman DA. The human microbiome: ecosystem resilience and health. *Nutr Rev* 70: S2-S9, 2012.
97. Roberts JD, Suckling CA, Peedle GY, Murphy JA, Dawkins TG, Roberts MG. An exploratory investigation of endotoxin levels in novice long distance triathletes, and the effects of a multi-strain probiotic/prebiotic, antioxidant intervention. *Nutrients* 8: 733, 2016.
98. Ryu D, Mouchiroud L, Andreux PA, Katsyuba E, Moullan N, Nicolet-Dit-Felix AA, Williams EG, Jha P, Lo Sasso G, Huzard D, Aebischer P, Sandi C, Rinsch C, Auwerx J. Urolithin A induces mitophagy and prolongs lifespan in *C. elegans* and increases muscle function in rodents. *Nat Med* 22: 879-888, 2016.
99. Schmidt TSB, Raes J, Bork P. The human gut microbiome: from association to modulation. *Cell* 172: 1198-1215, 2018.
100. Schubert K, Olde Damink SWM, von Bergen M, Schaap FG. Interactions between bile salts, gut microbiota, and hepatic innate immunity. *Immunol Rev* 279: 23-35, 2017.
101. Shen S, Kumar KP, Stanley D, Moore RJ, Van TTH, Wen SW, Hickey MJ, Wong CHY. Invariant natural killer T cells shape the gut microbiota and regulate neutrophil recruitment and function during intestinal inflammation. *Front Immunol* 9: 999, 2018.
102. Siddhart J, Chakrabarti A, Pannérec A, Karaz S, Morin-Rivron D, Masoodi M, Feige JN, Parkinson SJ. Aging and sarcopenia associate with specific interactions between gut microbes, serum biomarkers and host physiology in rats. *Aging* 9: 1698-1720, 2017.
103. Sket R, Debevec T, Kublik S, Schloter M, Schoeller A, Murovec B, Vogel Mikus K, Makuc D, Pecnik K, Plavec J, Mekjavic IB, Eiken O, Prevorsek Z, Stres B. Intestinal metagenomes and metabolomes in healthy young males: inactivity and hypoxia generated negative physiological symptoms precede microbial dysbiosis. *Front Physiol* 9: 198, 2018.
104. Sket R, Treichel N, Debevec T, Eiken O, Mekjavic I, Schloter M, Vital M, Chandler J, Tiedje JM, Murovec B, Prevorsek Z, Stres B. Hypoxia and inactivity related physiological changes (constipation, inflammation) are not reflected at the level of gut metabolites and butyrate producing microbial community: the PlanHab Study. *Front Physiol* 8: 250, 2017.
105. Sket R, Treichel N, Kublik S, Debevec T, Eiken O, Mekjavic I, Schloter M, Vital M, Chandler J, Tiedje JM, Murovec B, Prevorsek Z, Linar M, Stres B. Hypoxia and inactivity related physiological changes precede or take place in absence of sig-

- nificant rearrangements in bacterial community structure: the PlanHab randomized trial pilot study. *PLoS One* 12: e0188556, 2017.
106. Stewart LK, Flynn MG, Campbell WW, Craig BA, Robinson JP, McFarlin BK, Timmermann KL, Coen PM, Felker J, Talbert E. Influence of exercise training and age on CD14+ cell-surface expression of toll-like receptor 2 and 4. *Brain Behav Immun* 19: 389-397, 2005.
 107. Sung MM, Byrne NJ, Robertson IM, Kim TT, Samokhvalov V, Levasseur J, Soltys CL, Fung D, Tyreman N, Denou E, Jones KE, Seubert JM, Schertzer JD, Dyck JRB. Resveratrol improves exercise performance and skeletal muscle oxidative capacity in heart failure. *Am J Physiol Heart Circ Physiol* 312: H842-H853, 2017.
 108. Sun MF, Shen YQ. Dysbiosis of gut microbiota and microbial metabolites in Parkinson's Disease. *Ageing Res Rev* 45: 53-61, 2018.
 109. Tana C, Lauretani F, Ticinesi A, Prati B, Nouvenne A, Meschi T. Molecular and clinical issues about the risk of venous thromboembolism in older patients: a focus on Parkinson's disease and parkinsonism. *Int J Mol Sci* 19(5): E1299, 2018.
 110. Thaiss C, Levy M, Suez J, Elinav E. The interplay between the innate immune system and the microbiota. *Curr Opin Immunol* 26: 41-48, 2014.
 111. Thevaranjan N, Puchta A, Schulz C, Naidoo A, Szamosi JC, Verschoor CP, Loukov D, Schenck LP, Jury J, Foley KP, Schertzer JD, Larché MJ, Davidson DJ, Verdù EF, Surette MG, Bowdish DME. Age-associated microbial dysbiosis promotes intestinal permeability, systemic inflammation, and macrophage dysfunction. *Cell Host Microbe* 21: 455-466, 2017.
 112. Ticinesi A, Lauretani F, Milani C, Nouvenne A, Tana C, Del Rio D, Maggio M, Ventura M, Meschi T. Aging gut microbiota at the cross-road between nutrition, physical frailty and sarcopenia: is there a gut-muscle axis? *Nutrients* 9: E1303, 2017.
 113. Ticinesi A, Milani C, Guerra A, Allegri F, Lauretani F, Nouvenne A, Mancabelli L, Lugli GA, Turrone F, Duranti S, Mangifesta M, Viappiani A, Ferrario C, Dodi R, Dall'Asta M, Del Rio D, Ventura M, Meschi T. Understanding the gut-kidney axis in nephrolithiasis: an analysis of the gut microbiota composition and functionality of stone formers. *Gut* 67(12): 2097-2106, 2018.
 114. Ticinesi A, Milani C, Lauretani F, Nouvenne A, Mancabelli L, Lugli GA, Turrone F, Duranti S, Mangifesta M, Viappiani A, Ferrario C, Maggio M, Ventura M, Meschi T. Gut microbiota is associated with polypharmacy in elderly hospitalized patients. *Sci Rep* 7: 11102, 2017.
 115. Ticinesi A, Tana C, Nouvenne A, Prati B, Lauretani F, Meschi T. Gut microbiota, cognitive frailty and dementia in older individuals: a systematic review. *Clin Interv Aging* 13: 1497-1511, 2018.
 116. Ticinesi A, Tana C, Nouvenne A. The intestinal microbiome and its relevance for functionality in older persons. *Curr Opin Clin Nutr Metab Care* 22: 4-12, 2019.
 117. Tomkovich S, Jobin C. Microbiota and host immune responses: a love-hate relationship. *Immunology* 147: 1-10, 2015.
 118. Toohey JC, Townsend JR, Johnson SB, Toy AM, Vantrease WC, Bender D, Crimi CC, Stowers KL, Ruiz MD, Van Dusseldorp TA, Feito Y, Mangine GT. Effect of probiotic (*Bacillus subtilis*) supplementation during offseason resistance training in female division I athletes. *J Strength Cond Res* ahead of print, 2018. Doi: 10.1519/JSC0000000000002675.
 119. Townsend JR, Bender D, Vantrease WC, Sapp PA, Toy AM, Woods CA, Johnson KD. Effects of probiotic (*Bacillus subtilis* DE111) supplementation on immune function, hormonal status, and physical performance in division 1 baseball players. *Sports* 6: 70, 2018.
 120. Varian BJ, Goureshetti S, Poutahidis T, Lakritz JR, Levkovich T, Kwok C, Teliouis K, Ibrahim YM, Mirabal S, Erdman SE. Beneficial bacteria inhibit cachexia. *Oncotarget* 7: 11803-11816, 2016.
 121. Vazquez-Castellano JF, Serrano-Villar S, Jimenez-Hernandez N, Soto del Rio MD, Gayo S, Rojo D, Ferrer M, Barbas C, Moreno S, Estrada V, Rattei T, Latorre A, Moya A, Gosalbes MJ. Interplay between gut microbiota metabolism and inflammation in HIV infection. *ISME J* 12: 1964-1976, 2018.
 122. Vazquez-Castellanos JF, Serrano-Villar S, Latorre A, Artacho A, Ferrus ML, Madrid N, Vallejo A, Sainz T, Martinez-Botas J, Ferrando-Martinez S, Vera M, Dronza F, Leal M, Del Romero J, Moreno S, Estrada V, Gosalbes MJ, Moya A. Altered metabolism of gut microbiota contributes to chronic immune activation in HIV-infected individuals. *Mucosal Immunol* 8(4): 760-772, 2015.
 123. Ventura M, Turrone F, Canchaya C, Vaughan EE, O'Toole PW, van Sinderen D. Microbial diversity in the human intestine and novel insights from metagenomics. *Front Biosci* 14: 3214-3221, 2009.
 124. Vesterbacka J, Rivera J, Noyan K, Parera M, Neogi U, Calle M, Paredes R, Sonnerborg A, Noguera-Julian M, Nowak P. Richer gut microbiota with distinct metabolic profile in HIV infected elite controllers. *Sci Rep* 7: 6269, 2017.
 125. Walsch ME, Bhattacharya A, Sataranatarayan K, Qaisar R, Sloane L, Rahman MM, Kinter M, Van Remmen H. The histone deacetylase inhibitor butyrate improves metabolism and reduces muscle atrophy during aging. *Ageing Cell* 14: 957-970, 2015.
 126. Wilson D, Jackson T, Sapey E, Lord JM. Frailty and sarcopenia: the potential role of an aged immune system. *Ageing Res Rev* 36: 1-10, 2017.
 127. Yang Y, Shi Y, Wiklund P, Tan X, Wu N, Zhang X, Tikkanen O, Zhang C, Munukka E, Cheng S. The association between cardiorespiratory fitness and gut microbiota composition in premenopausal women. *Nutrients* 9: 792, 2017.
 128. Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, Magris M, Hidalgo G, Baldassano RN, Anokhin AP, Heath AC, Warner B, Reeder J, Kuczynski J, Caporaso JG, Lozupone CA, Lauber C, Clemente JC, Knights D, Knight R, Gordon JI. Human gut microbiome viewed across age and geography. *Nature* 486: 222-227, 2012.
 129. Yuan X, Xu S, Huang H, Liang J, Wu Y, Li C, Yuan H, Zhao X, Lai X, Hou S. Influence of excessive exercise on immunity, metabolism, and gut microbial diversity in an overtraining mice model. *Scand J Med Sci Sports* 28: 1541-1551, 2018.
 130. Zhang H, Luo XM. Control of commensal microbiota by the adaptive immune system. *Gut Microbes* 6: 156-160, 2015.
 131. Zhao X, Zhang Z, Hu B, Huang W, Yuan C, Zou L. Response of gut microbiota to metabolite changes induced by endurance exercise. *Front Microbiol* 9: 765, 2018.
 132. Zhernakova A, Kurilshikov A, Bonder MJ, Tigchelaar EF, Schirmer M, Vatanen T, Mujagic Z, Vila AV, Falony G, Vieira-Silva S, Wang J, Imhann F, Brandsma E, Jankipersadsing JA, Joossens M, Cenit MC, Deelen P, Swertz MA, LifeLines Cohort Study, Weersma RK, Feskens EJ, Netea MG, Gevers

- D, Jonkers D, Franke L, Aulchenko YS, Huttenhower C, Raes J, Hofker MH, Xavier MJ, Wijmenga C, Fu J. Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity. *Science* 352: 565-569, 2016.
133. Zilberman-Schapira G, Zmora N, Itav S, Bashiardes S, Elinav H, Elinav E. The gut microbiome in human immunodeficiency virus infection. *BMC Med* 14: 83, 2016.
134. Zmora N, Zilberman-Schapira G, Suez J, Mor U, Dori-Bachash M, Bashiardes S, Kotler E, Zur M, Regev-Lehavi D, Brik RBZ, Federici S, Cohen Y, Linevsky R, Rothschild D, Moor AE, Ben-Moshe S, Harmelin A, Itzkovitz S, Maharshak N, Shibolet O, Shapiro H, Pevsner-Fischer M, Sharon I, Halpern Z, Segal E, Elinav E. Personalized gut mucosal colonization resistance to empiric probiotics is associated with unique host and microbiome features. *Cell* 174: 1388-1405, 2018.